



**University of
Zurich**^{UZH}

**Zurich Open Repository and
Archive**

University of Zurich
University Library
Strickhofstrasse 39
CH-8057 Zurich
www.zora.uzh.ch

Year: 2009

Differential effects of yolk hormones on maternal and paternal contribution to parental care

Tschirren, Barbara ; Richner, Heinz

Abstract: In species with biparental care, a female gains fitness benefits from the joint reproductive investment of herself and her partner, but pays only the costs of her own care. Selection thus favours mechanisms that allow females to elicit a higher paternal investment from their partners. In oviparous species, the allocation of maternal yolk androgens to the eggs might represent such a female adaptation to sexually antagonistic selection. To test this hypothesis, we experimentally blocked the effects of maternal yolk androgens by an injection of the antiandrogen flutamide or a control substance in the eggs of great tits, *Parus major*. We subsequently manipulated the food demand of the brood in a brood size manipulation experiment, and recorded the parental feeding rates. The males' food-provisioning rates were not significantly influenced by the actions of maternal yolk androgens, whereas females adjusted their parental effort to androgen-mediated nestling signals, in particular in enlarged broods. These results show that female great tits do not exploit the male's contribution to parental care by allocating high concentrations of yolk androgens to their eggs. However, they indicate that variation in yolk androgen allocation among females has evolved through a process of coadaptation that matches maternal food provisioning and offspring demand.

DOI: <https://doi.org/10.1016/j.anbehav.2008.01.007>

Posted at the Zurich Open Repository and Archive, University of Zurich

ZORA URL: <https://doi.org/10.5167/uzh-76084>

Journal Article

Accepted Version

Originally published at:

Tschirren, Barbara; Richner, Heinz (2009). Differential effects of yolk hormones on maternal and paternal contribution to parental care. *Animal Behaviour*, 75(6):1989-1994.

DOI: <https://doi.org/10.1016/j.anbehav.2008.01.007>

**Differential effects of yolk hormones on maternal and paternal
contribution to parental care**

BARBARA TSCHIRREN^{1, 2} & HEINZ RICHNER²

¹Centre for the Integrative Study of Animal Behaviour, Macquarie University

²Evolutionary Ecology Group, University of Bern

*Author for correspondence:

Barbara Tschirren, Centre for the Integrative Study of Animal Behaviour,
Macquarie University, Sydney, NSW 2109, Australia; e-mail:
barbara.tschirren@unsw.edu.au

Heinz Richner, Evolutionary Ecology Group, Zoological Institute, University of
Bern, Baltzerstrasse 6, 3012 Bern, Switzerland; e-mail:
heinz.richner@esh.unibe.ch

Running headline:

Tschirren & Richner: Yolk hormones and parental investment

Word count: 5057 words

In species with biparental care, a female gains fitness benefits from the joint reproductive investment of herself and her partner, but pays only the costs of her own care. Selection thus favours mechanisms that allow females to elicit a higher paternal investment from her partner. In oviparous species, the allocation of maternal yolk androgens into the eggs might represent such a female adaptation to sexually antagonistic selection. To test this hypothesis, we experimentally blocked the effects of maternal yolk androgens by an injection of the anti-androgen flutamide or a control substance in the eggs of great tits (*Parus major*). After hatching, we subsequently manipulated the food demand of the brood in a brood size manipulation experiment, and recorded the parental feeding rates. We found that the males' food provisioning rates were not significantly influenced by the actions of maternal yolk androgens, while females adjusted their parental effort to androgen-mediated nestling signals, in particular in enlarged broods. These results show that female great tits do not exploit the male's contribution to parental care by allocating high concentrations of yolk androgens into their eggs. It however indicates, that variation in yolk androgen allocation among females has evolved through a process of coadaptation that matches maternal food provisioning and offspring demand.

Keywords: coadaptation, food provisioning, great tit, maternal effects, maternal yolk androgens, parental investment, parent-offspring communication, *Parus major*, sexual conflict

49 In sexually reproducing species, a conflict over parental investment occurs
50 because each parent gains fitness benefits from the joint reproductive
51 investment of both partners, but pays only the costs of its own care. Selection
52 will thus favour adaptations that successfully manipulate the genetically
53 unrelated partner into elevating its parental investment, which, as a
54 consequence, will allow the manipulator to reduce its own reproductive
55 contribution (Trivers 1972; Houston & Davies 1985; Lessells 1999; McNamara
56 et al. 1999; Arnqvist & Rowe 2005; Houston et al. 2005).

57 In blue tits (*Cyanistes caeruleus*), for example, a sexual conflict
58 over the degree of hatching asynchrony occurs (Slagsvold et al. 1994, 1995).
59 Females pay relatively lower costs of parental care in terms of future survival
60 when raising even-aged nestlings, while males survive better when caring for
61 an asynchronous brood (Slagsvold et al. 1994, 1995). While the total cost of
62 parental care is similar for synchronous and asynchronous broods, the
63 different hatching patterns thus alter the relative reproductive burden on the
64 two sexes (Slagsvold et al. 1994, 1995). Because only the female incubates in
65 this species, she can control the degree of hatching asynchrony by varying
66 the start of incubation, and is thus likely to win this conflict (Slagsvold et al.
67 1994, 1995).

68 Not only the start of incubation, but any reproductive behaviour that
69 is under female control alone, has the potential to mediate sexual conflicts
70 over reproductive decisions in favour of the female's own interest. In birds
71 (Schwabl 1993; Groothuis et al. 2005), fish (McCormick 1999), and reptiles
72 (Bowden et al. 2000; Lovern & Wade 2003), for example, females are known
73 to deposit androgens and other hormones into their eggs. The amount of yolk

74 hormones varies however greatly, not only among species, but also within and
75 among clutches of the same species and even among clutches of the same
76 female. Variation within clutches, i.e. increasing or decreasing hormone
77 concentrations with laying order, might aim at mitigating the competitive
78 asynchrony among siblings caused by asynchronous hatching (e.g. Eising et
79 al. 2001; Pilz et al. 2003) or enhance these effects to facilitate brood reduction
80 when food is limited (Schwabl et al. 1997). However, although variation in the
81 amount of yolk hormones deposited *among* clutches of different females is
82 typically even larger (e.g. Reed & Vleck 2001; Groothuis & Schwabl 2002; Pilz
83 et al. 2003; Tschirren et al. 2004), the mechanisms driving this variation are
84 as yet poorly understood (Groothuis et al. 2005, Müller et al. 2007b).

85 Experimental studies on a number of bird species have shown that
86 yolk androgens deposited by the mother can have profound effects on the
87 development, morphology and food acquisition behaviour of nestlings
88 (reviewed in Groothuis et al. 2005). Young birds originating from eggs with
89 experimentally high yolk androgens concentrations showed, for example, a
90 higher begging effort (Schwabl, 1996; Eising & Groothuis 2003; von
91 Engelhardt et al. 2006; but see Pilz et al. 2004; Boncoraglio et al. 2006) and a
92 higher growth rate (Schwabl, 1996; Eising et al. 2001; Tschirren et al. 2005)
93 than their unmanipulated siblings. Further, maternal yolk androgens can
94 promote the development of the *musculus complexus*, a neck muscle involved
95 in food acquisition (Lipar & Ketterson 2000), and beak flange size, which
96 might stimulate parental food provisioning (Müller et al. 2007b). Yolk
97 hormones can thus influence a variety of physiological, behavioural and

98 morphological nestling traits, all of which may affect parental feeding
99 decisions.

100 If males are especially sensitive to such hormone-dependent nestling
101 signals, then the deposition of yolk androgens by the female could represent a
102 sexually antagonistic adaptation that aims at exploiting the male's contribution
103 to parental care, as recently suggested by Michl et al. (2005), Moreno-Rueda
104 (2007) and Müller et al. (2007a). To test this, we experimentally inhibited the
105 effects of maternal yolk androgens by injecting flutamide, an androgen
106 receptor inhibitor, or a control substance into the eggs of free-living great tits
107 (*Parus major*), and filmed the food provisioning behaviour of the parents to
108 measure their investment.

109 Flutamide directly competes with testosterone and testosterone-
110 metabolites for binding to androgen receptors (Simard et al. 1986), and
111 previous work on chicken (*Gallus gallus*; Burke 1996), red-winged blackbirds
112 (Lipar & Ketterson 2000) and black-headed gulls (Müller et al. 2005) has
113 confirmed its anti-androgenic actions when injected *in ovo*. If the deposition of
114 yolk androgens into the eggs is a sexually antagonistic adaptation of the
115 female to increase the male's contribution to parental care, we predict 1)
116 higher feeding rates of the male in control-injected compared to flutamide-
117 injected (i.e. androgen-blocked) broods, and 2) that the effect of yolk
118 androgens on the parental food provisioning behaviour will be most
119 pronounced when the nestlings' degree of hunger, and thereby the cost of
120 rearing the brood, is increased (i.e. in experimentally enlarged broods).

121

122

METHODS

Study Site, Species and Flutamide Injection

The experiment was performed in 2003 in a great tit population (*Parus major*) breeding in nest boxes in a forest near Bern, Switzerland ("Forst", 46°54'N 7°17'E / 46°57'N 7°21'E). Great tits are socially monogamous passerines that rear one, or in a few cases two broods per year. Only females build the nest and incubate the eggs but both parents feed the young during the nestling stage (see e.g. Tschirren et al. 2005a for a typical division of work between the sexes).

All nest boxes were cleaned before the start of the breeding season (February) to remove nest-based ectoparasites, which can influence yolk androgen deposition (Tschirren et al. 2004). From the beginning of the breeding season (April) onwards, we regularly visited nest boxes to determine the start of nest building and egg laying. After clutch completion, we injected all eggs of a clutch with either 0.0319µmol of the anti-androgen Flutamide (Fluka, Switzerland) dissolved in 5µl ethanol (70%) or with 5µl ethanol (70%) as a control (see Lipar & Ketterson 2000 for details). The injected dose of flutamide was based on the amounts used in Burke (1996) and Lipar & Ketterson (2000), adjusted for an average yolk mass of 352mg in our study population. The injections were done in the field using a 25µl syringe (Hamilton 702LT) and a 25 G needle (see Tschirren et al. 2005b for details). During the injection procedure, the eggs were illuminated from beneath using a cold light source (Intralux 4000, Volpi, Switzerland) to ensure that the tip of the needle penetrated the yolk membrane. The hole in the eggshell was sealed by applying a small drop of tissue adhesive (Vetseal, B. Braun

Medical, Switzerland). The average hatching success of the eggs was 77.2%.
It did not significantly differ between flutamide-injected and control eggs ($\chi^2 = 0.45$, $P = 0.502$, $N = 534$).

Brood Size Manipulation

One day after hatching, we created broods of reduced (-2 nestlings compared to original clutch size) and enlarged size (+ 2 nestlings compared to original clutch size) by partially exchanging randomly chosen nestlings among nests with the same flutamide treatment, the same hatching date and a similar clutch size. After the manipulation, both enlarged and reduced nests contained own and foster nestlings of the same flutamide-treatment group, and the manipulated brood sizes remained within the range of natural variation observed in our study population (5–12 nestlings / brood). This manipulation allowed us to assess the investment of the parents in response to an elevated or reduced food demand of the brood, and the interaction effects between the food demand of the brood and the effects of maternal yolk androgens (i.e. a 2x2 design).

The original clutch size did not differ significantly between treatment groups (brood size manipulation, $F_{1, 46} = 0.01$, $P = 0.941$; flutamide treatment, $F_{1, 46} = 0.81$, $p = 0.373$; interaction, $F_{1, 45} = 0.07$, $P = 0.800$). After the manipulation, the brood size was significantly larger in enlarged compared to reduced broods (brood size manipulation, $F_{1, 46} = 187.24$, $P < 0.001$; $R^2 = 0.80$), but did not differ significantly between the flutamide-treatment groups ($F_{1, 46} = 0.42$, $P = 0.520$; interaction, $F_{1, 45} = 0.08$, $P = 0.776$).

Parental Food Provisioning

Ten days post-hatching, when the nestlings' food demand is highest (Gebhardt-Henrich 1990; Keller & Van Noordwijk 1994), we installed a video camera sensitive to infrared and equipped with an infrared lamp in the nest box to assess the parental investment in response to the flutamide treatment and the brood size manipulation. Video recording started 30 minutes after camera installation and the food provisioning behaviour of the parents was filmed during the following 100 minutes. After the filming, nestlings were weighted and ringed with individually numbered aluminium rings.

A total of 49 broods (11 enlarged, control-injected broods, 14 reduced, control-injected broods, 11 enlarged, flutamide-injected broods, and 13 reduced, flutamide-injected broods) were filmed. The analysis of the recordings was performed blindly with respect to the treatment of the nest.

Males and females can be visually identified on the videos. Only visits during which the male or female parent delivered food to at least one nestling were counted as a feeding visit. In addition to the number of parental feeding visits, we also classified the prey size brought to the nestlings as small, intermediate or large (Kölliker et al. 1998). The food quantity delivered to the nestlings was then calculated as the product of feedings per hour and mean prey size.

Because the analyses of feeding visits and food quantity delivered to the brood were qualitatively very similar (see also Moreno et al. 1995; Neuenschwander et al. 2003) only the results of the former are presented here.

The disturbance of the nest during the installation of the camera (performed within < 5 minutes) and the filming within the nest boxes (method

established in previous years; see e.g. Tschirren et al. 2005a) did not have a negative effect on the feeding behaviour of the parents. Both parents fed the nestlings on a regular basis during the filming in all experimental nests, and no nest desertion or nestling mortality was observed during the filming. The video recording or the disturbance of the nests during the manipulations did not have any observable adverse short- or long-term effects on the nestlings, and their weight (mean: $15.77 \pm 0.10\text{g}$) was similar to the weight of great tit nestlings in our study population in previous years (range of mean weights between 1997 and 2002: 14.49g–16.21g, depending on food availability).

Statistical Analyses

The flutamide treatment and the brood size manipulation influenced the feeding behaviour of males and females differently (see Results). We thus analysed the feeding behaviour of mothers and fathers separately. Maternal and paternal feeding visits were analysed with ANCOVAs including the flutamide treatment, the brood size manipulation, and their interaction as fixed effects and nestling body mass at filming as a covariate. The interaction was removed from the final model if non-significant. For the analysis of differences between flutamide-injected and control-injected broods within a given brood size manipulation group we used least-square mean contrasts.

All tests were two-tailed with a significance level of $P \leq 0.05$. Residuals of the models were tested for normality using Shapiro-Wilk tests and homoscedasticity using Bartlett tests to ensure that the assumptions for parametric testing were fulfilled. Means ± 1 S.E. are presented in the results

and figures. Statistical analyses were performed using JMP IN 5.1 (Sall & Lehmann 1996).

RESULTS

The flutamide treatment and the brood size manipulation influenced the feeding behaviour of the two parents differently (three-way interaction: parental sex x brood size manipulation x flutamide treatment, $F_{1,44} = 6.08$, $P = 0.018$). We therefore analysed the feeding behaviour of mothers and fathers separately.

Paternal Food Provisioning

Contrary to our prediction, we found no significant effect of the flutamide treatment ($F_{1,45} = 0.17$, $P = 0.685$) and no significant interaction effect of the flutamide treatment and the brood size manipulation ($F_{1,44} < 0.01$, $P = 0.993$) on the males' feeding rates (Fig. 1a). Independent of the flutamide treatment of the young, males raising an enlarged brood fed at significantly higher rates than males raising a reduced brood ($F_{1,45} = 4.87$, $P = 0.033$, Fig. 1a).

Male feeding rates were not significantly influenced by nestling body mass at filming ($F_{1,45} = 2.61$, $P = 0.114$). Further, no effect of the brood size manipulation on the males' feeding visits *per nestling* were found ($F_{1,45} = 1.95$, $P = 0.170$), showing that males increased their feeding effort in enlarged broods to keep the food provisioning per nestling constant. Male feeding rates were not significantly influenced by the date of filming ($F_{1,41} = 2.19$, $P = 0.147$), the time of filming ($F_{1,41} < 0.01$, $P = 0.932$), the original clutch size ($F_{1,41} = 0.31$, $P = 0.578$), or the partner's feeding rate ($F_{1,41} = 0.174$, $P = 0.679$).

Maternal Food Provisioning

We found a significant interaction effect of the flutamide treatment and the brood size manipulation on the mother's feeding visits ($F_{1, 44} = 8.65$, $P = 0.005$, Fig. 1b). When the food demand of the brood was low (i.e. in reduced broods), mothers did not feed control-injected and flutamide-injected nestlings at significantly different rates (contrast: $F_{1, 44} = 0.04$, $P = 0.851$), whereas when the food demand of the brood was high (i.e. in enlarged broods), mothers of control-injected broods showed significantly higher feeding rates compared to mothers of flutamide-injected broods (contrast: $F_{1, 44} = 17.15$, $P < 0.001$). Similarly, within the control-injected group, mothers feeding an enlarged brood showed significantly higher feeding rates than mothers feeding a reduced brood (contrast $F_{1, 44} = 21.50$, $P < 0.001$), while in the flutamide-injected group the difference in the females' food provisioning behaviour between enlarged and reduced broods was not statistically significant (contrast $F_{1, 44} = 0.46$, $P = 0.499$). Thus, females did not adjust their food provisioning to the higher food demand of enlarged broods when the actions of maternal yolk hormones were blocked. Similar results were found when analysing the females' feeding visits per nestling instead of the total feeding visits per brood (interaction brood size manipulation x hormone treatment: $F_{1, 44} = 4.30$, $P = 0.044$), indicating that females overcompensated in response to the brood size manipulation.

Female feeding rates significantly increased during the breeding season ($F_{1, 43} = 4.27$, $P = 0.045$) and tended to decrease with increasing nestling body mass ($F_{1, 43} = 3.03$, $P = 0.089$). However, including these covariates into the analysis did not change the interaction effect between the

brood size manipulation and the hormone treatment on female feeding rates (interaction: $F_{1, 43} = 10.28$, $P = 0.003$). Female feeding rates were not significantly influenced by the time of video recording ($F_{1, 40} = 0.13$, $P = 0.723$), the original clutch size ($F_{1, 40} = 0.17$, $P = 0.681$), or the partner's feeding rate ($F_{1, 40} = 0.13$, $P = 0.718$).

DISCUSSION

In this study we experimentally investigated if maternal yolk androgens represent a mechanism by which females can manipulate their partner's investment in parental care, as recently suggested by Michl et al. (2005), Moreno-Rueda (2007) and Müller et al. (2007a). Counter to the predictions of this hypothesis, we found that males adjusted their feeding effort solely to the number of nestlings present in the nest, irrespective of the flutamide-treatment of the young. Likewise, in control-injected broods, females increased their feeding visits when caring for an enlarged brood. However, when the actions of maternal yolk androgens were experimentally inhibited, (i.e. in the flutamide-injected group), an increased food demand of the brood did not lead to a higher maternal investment. Thereby these results indicate that yolk androgens allow nestlings to elicit higher feeding rates from their mother, especially when their food demand is increased (i.e. when environmental conditions get unfavourable; see also Pilz et al. 2004).

Our finding that yolk hormones have an effect on maternal, but not on paternal investment, suggests that selection may have reduced the males' susceptibility to yolk androgen-mediated nestling signals to resist the exploitation of their parental effort by the female (see also Müller et al. 2007a).

Indeed, evidence that males ignore certain nestling displays has been found in canaries, where males adjust their food provisioning to the chicks' begging posture only, while females respond to a variety of offspring signals (Kilner 2002b). Similarly, male great tits adjust their feeding behaviour to visual displays, while females integrate visual as well as vocal nestling signals (Kilner 2002a).

While males were not susceptible to the effects of yolk androgens, females did adjust their feeding behaviour in response to the flutamide treatment, especially when feeding an enlarged brood. This shows that maternal yolk hormones play an important role in regulating mother-offspring interactions. Indeed, Kölliker et al. (2000) showed by means of a partial cross-fostering experiment, that the level of food solicitation by the nestlings is largely dependent on their nest of origin, and that the female's increase in food provisioning in response to an increased demand was positively correlated to the natural levels of begging intensity of her offspring. No such correlation was found in the paternal line. Although the role of (additive) genetic variation cannot be excluded (Kölliker et al. 2000), our finding supports the idea that this correlation between offspring demand and maternal response may at least partly be mediated or enhanced by maternal effects (see also Kölliker et al. 2000; Kölliker et al. 2005).

The goal of this study was to investigate the effects of yolk hormones on parental feeding investment, which does not require a direct measurement of the numerous potential pathways that could mediate such effects. However, now we know that yolk hormones do indeed affect maternal investment, and that this effect is independent of nestling body mass, elucidating these

proximate mechanisms is an obvious next step. In particular, to test whether males specifically ignore yolk androgen-dependent nestling signals to avoid exploitation of parental care by the female, we will need to experimentally investigate the effects of yolk hormones on as many different aspects of offspring behaviour and morphology as possible, and to test for sex-specific sensitivities to these traits.

In conclusion, our study shows that female great tits do not exploit the male's contribution to parental care by differentially allocating yolk androgens into the eggs. However, the females' own adjustment of parental effort in response to yolk androgen-mediated nestling signals indicates that maternal hormones do play an important role in mother-offspring communication, and that variation in the deposition of yolk androgens between females may have evolved to optimally match maternal supply and offspring demand, as predicted by Kölliker et al. (2005). This maternal sensitivity to the actions of yolk hormones might ensure a prime start for offspring, and particularly for those of high reproductive value (see e.g. Gil et al. 1999, 2004; Tanvez et al. 2004; Loyau et al. 2007), even if environmental conditions get unfavourable. Interestingly, the mother's sensitivity to her offspring's need also makes her vulnerable to exploitation by her nestlings (Trivers 1974; Godfray 1995). However, our finding that females did, unlike males, maintain their sensitivity to yolk androgen-mediated nestling signals suggests that the costs of ignoring such signals are sex-specific, and that in females they do not exceed the benefits. Although more research is necessary to gain a full understanding of the adaptive value of variation in yolk hormone deposition among females, and the covariance between provisioning and soliciting behaviour caused by

genetic variation in maternal hormone levels, our study indicates that maternal
yolk hormones can play an important role in shaping the coadaptation among
mothers and their young.

ACKNOWLEDGEMENTS

We are grateful to Verena Saladin, Oliver Otti and Sibylle Aschwanden for
field assistance, Fabian Schibler for help with analysing videos, and Erik
Postma and Mathias Kölliker for discussion. The comments of the editor D. Gil
and anonymous reviewers greatly improved previous versions of the
manuscript. This work was financially supported by the Swiss National
Science Foundation (research grant no. 102017 to H.R.) and the Australian
Research Council (APD fellowship and research grant no. DP0879313 to
B.T.). It was conducted under a license provided by the Ethical Committee of
the Office of Agriculture of the Canton of Bern, Switzerland (No 05/03, 2003).

REFERENCES

Arnqvist, G. & Rowe, L. 2005. Parental care and sexual conflict. In: *Sexual
conflict* (Ed. by G. Arnqvist & L. Rowe), pp. 156-178. Princeton, New Jersey:
Princeton University Press.

371 **Boncoraglio, G, Rubolini, D, Romano, M., Martinelli, R. & Saino, N.** 2006.
 372 Effects of elevated yolk androgens on perinatal begging behavior in yellow-
 373 legged gull (*Larus michahellis*) chicks. *Hormones and Behavior*, **50**, 442-447.

374 **Bowden, R. M., Ewert, M. A. & Nelson, C. E.** 2000. Environmental sex
 375 determination in a reptile varies seasonally and with yolk hormones.
 376 *Proceedings of the Royal Society of London, Series B*, **267**, 1745-1749.

377 **Burke, W. H.** 1996. Effects of an *in ovo* injection of an anti-androgen on
 378 embryonic and posthatching growth of broiler chicks. *Poultry Science*, **75**,
 379 648-655.

380 **Eising, C. M. & Groothuis, T. G. G.** 2003. Yolk androgens and begging
 381 behaviour in black-headed gull chicks: an experimental field study. *Animal*
 382 *Behaviour*, **66**, 1027-1034.

383 **Eising, C. M., Eikenaar, C., Schwabl, H. & Groothuis, T. G. G.** 2001.
 384 Maternal androgens in black-headed gull (*Larus ridibundus*) eggs:
 385 consequences for chick development. *Proceedings of the Royal Society of*
 386 *London, Series B*, **268**, 839-846.

387 **Gebhardt-Henrich, S. G.** 1990. Temporal and spatial variation in food
 388 availability and its effect on fledgling size in the great tit. In: *Population biology*
 389 *of passerine birds: an integrated approach* (Ed. by J. Blondel, A. Gosler, J. D.
 390 Lebreton & R. McCleery), pp. 175-186. Berlin: Springer.

391 **Gil, D., Graves, J., Hazon, N. & Wells, A.** 1999. Male attractiveness and
 392 differential testosterone investment in zebra finch eggs. *Science*, **286**, 126-
 393 128.

394 **Gil, D., Leboucher, G., Lecroix, A., Cue, R. & Kreutzer, M.** 2004. Female
 395 canaries produce eggs with greater amount of testosterone when exposed to
 396 preferred male song. *Hormones and Behavior*, **45**, 64-70.

397 **Godfray, H. C. J.** 1995. Evolutionary theory of parent-offspring conflict.
 398 *Nature*, **376**, 133-138.

399 **Groothuis, T. G. & Schwabl, H.** 2002. Determinants of within- and among-
 400 clutch variation in levels of maternal hormones in black-headed gull eggs.
 401 *Functional Ecology*, **16**, 281-289.

402 **Groothuis, T. G. G., Müller, W., von Engelhardt, N., Carere, C. & Eising,**
 403 **C.** 2005. Maternal hormones as a tool to adjust offspring phenotype in avian
 404 species. *Neuroscience and Biobehavioral Reviews*, **29**, 329-352.

405 **Houston, A. I. & Davies, N. B.** 1985. The evolution of cooperation and life
 406 history in the dunnoek *Prunella modularis*. In: *Behavioural Ecology* (Ed. by R.
 407 M. Sibly & R. H. Smith), pp. 471-487. Oxford: Blackwell Scientific Publications.

408 **Houston, A. I., Székely, T. & McNamara, J. M.** 2005. Conflict between
 409 parents over care. *Trends in Ecology & Evolution*, **20**, 33-38.

410 **Keller, L. F. & Van Noordwijk, A. J.** 1994. Effects of local environmental-
 411 conditions on nestling growth in the great tit *Parus major* L. *Ardea*, **82**, 349-
 412 362.

413 **Kilner, R.** 2002a. The evolution of complex begging displays. In: *The*
 414 *evolution of begging, competition, cooperation and communication*. (Ed. by J.
 415 Wright & M. L. Leonard), pp. 87-106. Dordrecht: Kluwer Academic Publisher.

416 **Kilner, R. M.** 2002b. Sex differences in canary (*Serinus canaria*) provisioning
 417 rules. *Behavioral Ecology and Sociobiology*, **52**, 400-407.

418 **Kölliker, M., Richner, H., Werner, I. & Heeb, P.** 1998. Begging signals and
 419 biparental care: nestling choice between parental feeding locations. *Animal*
 420 *Behaviour*, **55**, 215-222.

421 **Kölliker, M., Brinkhof, M. W. G., Heeb, P., Fitze, P. S. & Richner, H.** 2000.
 422 The quantitative genetic basis of offspring solicitation and parental response
 423 in a passerine bird with biparental care. *Proceedings of the Royal Society of*
 424 *London, Series B*, **267**, 2127-2132.

425 **Kölliker, M., Brodie, E. D. & Moore, A. J.** 2005. The coadaptation of parental
 426 supply and offspring demand. *American Naturalist*, **166**, 506-516.

427 **Lessells, C. M.** 1999. Sexual conflict in animals. In: *Levels of selection in*
 428 *evolution* (Ed. by L. Keller), pp. 75-99. Princeton, New Jersey: Princeton
 429 University Press.

430 **Lipar, J. L. & Ketterson, E. D.** 2000. Maternally derived yolk testosterone
 431 enhances the development of the hatching muscle in the red-winged blackbird
 432 *Agelaius phoeniceus*. *Proceedings of the Royal Society, Series B*, **267**, 2005-
 433 2010.

434 **Lovern, M. B. & Wade, J.** 2003. Yolk testosterone varies with sex in eggs of
 435 the lizard, *Anolis carolinensis*. *Journal of Experimental Zoology, Part A*, **295A**,
 436 206-210.

437 **Loyau, A., Saint Jalme, M., Mauget, R. & Sorci, G.** 2007. Male
 438 attractiveness affects the investment of maternal resources into the eggs in
 439 peafowl (*Pavo cristatus*). *Behavioral Ecology and Sociobiology*, **61**, 1043-
 440 1052.

441 **McCormick, M. I.** 1999. Experimental test of the effect of maternal hormones
 442 on larval quality of a coral reef fish. *Oecologia*, **118**, 412-422.

443 **McNamara, J. M., Gasson, C. E. & Houston, A. I.** 1999. Incorporating rules
 444 for responding into evolutionary games. *Nature*, **401**, 368-371.

445 **Michl, G., Török, J., Péczely, P., Garamszegi, L. Z. & Schwabl, H.** 2005.
 446 Female collared flycatchers adjust yolk testosterone to male age, but not to
 447 attractiveness. *Behavioral Ecology*, **16**, 383-388.

448 **Moreno, J., Cowie, R. J., Sanz, J. J. & Williams, R. S. R.** 1995. Differential
 449 response by males and females to brood manipulations in the pied flycatcher -
 450 energy-expenditure and nestling diet. *Journal of Animal Ecology*, **64**, 721-732.

451 **Moreno-Rueda, G.** 2007. Yolk androgen deposition as a female tactic to
 452 manipulate paternal contribution. *Behavioral Ecology*, **18**, 496-498.

453 **Müller, W., Groothuis, T. G. G., Eising, C. M. & Dijkstra, C.** 2005. An
 454 experimental study on the causes of sex-biased mortality in the black-headed
 455 gull - the possible role of testosterone. *Journal of Animal Ecology*, **74**, 735-
 456 741.

457 **Müller, W., Lessells, C. M., Korsten, P. & von Engelhardt, N.** 2007a.
 458 Manipulative signals in family conflict? On the function of maternal yolk
 459 hormones in birds. *American Naturalist*, **169**, E84-96.

460 **Müller, W., Deptuch, K., López-Rull, I. & Gil, D.** 2007b. Elevated yolk
 461 androgen levels benefit offspring development in a between-clutch context.
 462 *Behavioral Ecology* **18**, 929-936.

463 **Neuenschwander, S., Brinkhof, M. W. G., Kölliker, M. & Richner, H.** 2003.
 464 Brood size, sibling competition, and the cost of begging in great tits (*Parus*
 465 *major*). *Behavioral Ecology*, **14**, 457-462.

466 **Pilz, K. M., Smith, H. G., Sandell, M. I. & Schwabl, H.** 2003. Interfemale
 467 variation in egg yolk androgen allocation in the European starling: do high-
 468 quality females invest more? *Animal Behaviour*, **65**, 841-850.

469 **Pilz, K. M., Quiroga, M., Schwabl, H. & Adkins-Regan, E.** 2004. European
 470 starling chicks benefit from high yolk testosterone levels during a drought
 471 year. *Hormones and Behavior*, **46**, 179-192.

472 **Reed, W. L. & Vleck, C. M.** 2001. Functional significance of variation in egg-
 473 yolk androgens in the American coot. *Oecologia*, **128**, 164-171.

474 **Sall, J. & Lehmann, A.** 1996. JMP Start Statistics. New York: Duxbury
 475 Press.

476 **Schwabl, H.** 1993. Yolk is a source of maternal testosterone for developing
 477 birds. *Proceedings of the National Academy of Sciences of the United States*
 478 *of America*, **90**, 11446-11450.

479 **Schwabl, H.** 1996. Maternal testosterone in the avian egg enhances postnatal
 480 growth. *Comparative Biochemistry and Physiology A*, **114**, 271-276.

481 **Schwabl, H., Mock, D. W. & Gieg, J. A.** 1997. A hormonal mechanism for
 482 parental favouritism. *Nature*, **386**, 231-231.

483 **Simard, J., Luthy, I., Guay, J., Belanger, A. & Labrie, F.** 1986.
 484 Characteristics of interaction of the antiandrogen flutamide with the androgen
 485 receptor in various target tissues. *Molecular and Cellular Endocrinology*, **44**,
 486 261-270.

487 **Slagsvold, T., Amundsen, T. & Dale, S.** 1994. Selection by sexual conflict
 488 for evenly spaced offspring in blue tits. *Nature*, **370**, 136-138.

489 **Slagsvold, T., Amundsen, T. & Dale, S.** 1995. Costs and benefits of
 490 hatching asynchrony in blue tits *Parus caeruleus*. *Journal of Animal Ecology*,
 491 **64**, 563-578.

492 **Tanvez, A., Béguin, N., Chastel, O., Lecroix, A. & Leboucher, G.** 2004.
 493 Sexually attractive phrases increase yolk androgens deposition in Canaries
 494 (*Serinus canaria*). *General and Comparative Endocrinology*, **138**, 113-120.

495 **Trivers, R. L.** 1972. Parental investment and sexual selection. In: *Sexual*
 496 *selection and the descent of man* (Ed. by B. Campbell), pp. 136–179.
 497 Chicago: Aldine.

498 **Trivers, R. L.** 1974. Parent-offspring conflict. *American Zoologist*, **14**, 249-
 499 264.

500 **Tschirren, B., Richner, H. & Schwabl, H.** 2004. Ectoparasite-modulated
 501 deposition of maternal androgens in great tit eggs. *Proceedings of the Royal*
 502 *Society of London, Series B*, **271**, 1371-1375.

503 **Tschirren, B., Fitze, P. S. & Richner, H.** 2005a. Carotenoid-based nestling
 504 colouration and parental favouritism in the great tit. *Oecologia*, **143**, 477-482.

505 **Tschirren, B., Saladin, V., Fitze, P. S., Schwabl, H. & Richner, H.** 2005b.
 506 Maternal yolk testosterone does not modulate parasite susceptibility or
 507 immune function in great tit nestlings. *Journal of Animal Ecology*, **74**, 675-682.

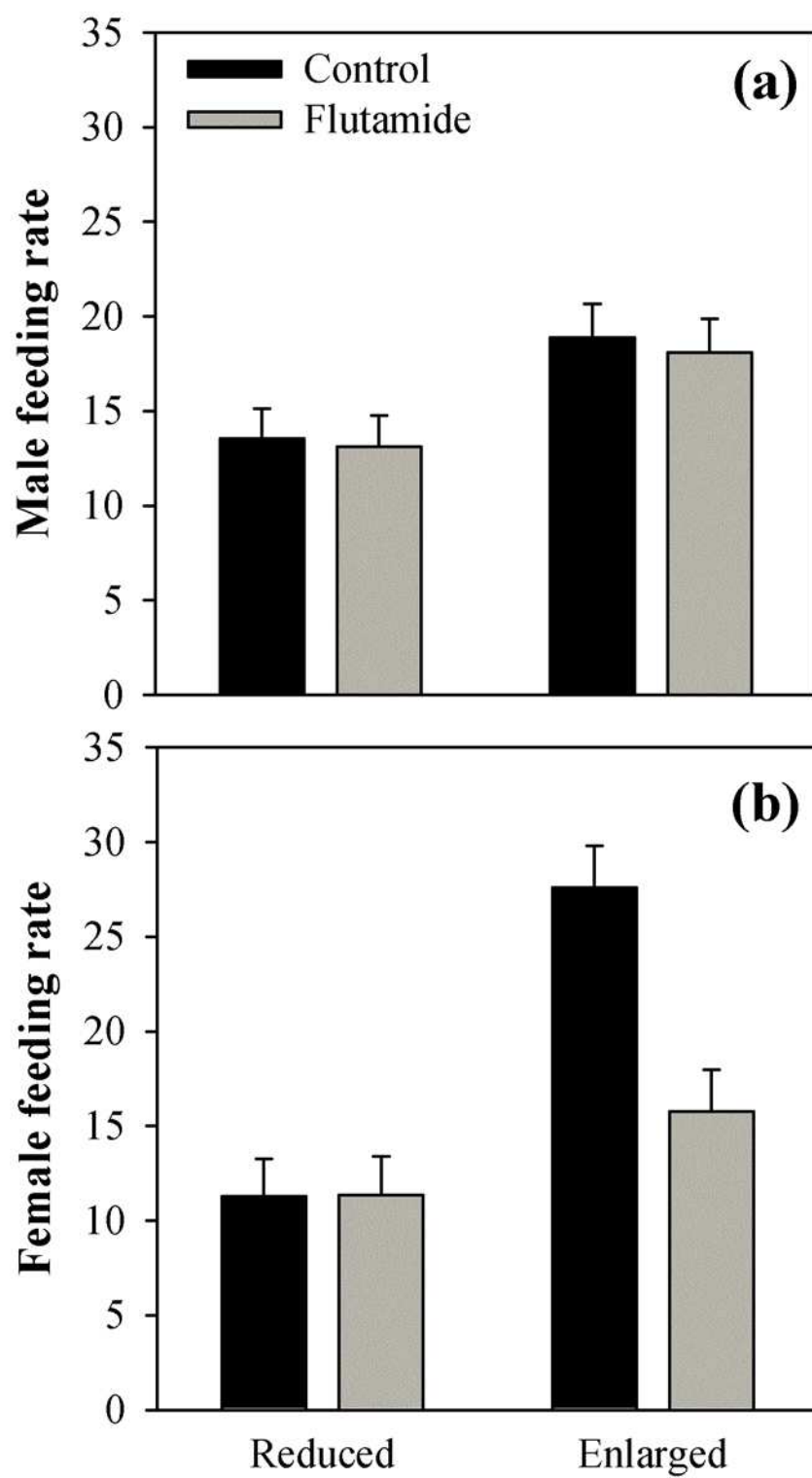
508 **von Engelhardt, N., Carere, C., Dijkstra, C. & Groothuis, T.G.G.** 2006. Sex-
 509 specific effects of yolk testosterone on survival, begging and growth of zebra
 510 finches *Proceedings of the Royal Society of London, Series B*, **273**, 65-70.

511

FIGURE LEGEND

Figure 1

Feeding visits per hour of a) male, and b) female great tits in relation to the flutamide treatment of the nestlings (nestlings originating from flutamide- or control-injected eggs) and the manipulated brood size (feeding an enlarged or reduced brood). Least square means + 1SE are shown.



521

522 **Fig. 1**